Supporting Information

Protease Activity Analysis: A Toolkit for Analyzing Enzyme Activity Data

Ava P. Soleimany,* $,\dagger,\dagger,\P,\#$ Carmen Martin-Alonso, $,\dagger,\#$ Melodi Anahtar,,# Cathy S. Wang,, and Sangeeta N. Bhatia, $,*,\dagger,\parallel,\perp$

†Harvard-MIT Division of Health Sciences and Technology, MIT, Cambridge, MA

‡Program in Biophysics, Harvard University, Boston, MA

¶Microsoft Research New England, Cambridge, MA

§Department of Biological Engineering, MIT, Cambridge, MA

∥Department of Electrical Engineering and Computer Science, MIT, Cambridge, MA

⊥Howard Hughes Medical Institute, Cambridge, MA

#Equal contribution.

*E-mail: avasoleimany@microsoft.com; sbhatia@mit.edu

Supporting Information (3 pages)

Code availability; Figure S1: Empirical validation of predictions for substrate ranking; Figure S2: Automated analysis of protease activity data from the literature.

Code availability

All source code for the PAA package and step-by-step tutorials can be found at https://github.com/apsoleimany/protease_activity_analysis.

Additional Results

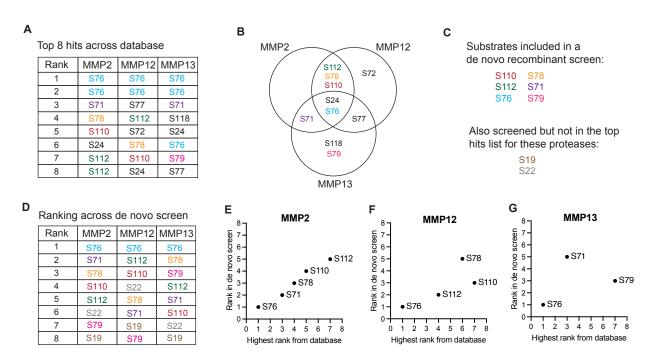


Figure S1: **PAA** predicts substrate rankings that are robust empirically. (A) PAA's database was queried to obtain the top substrate hits for three MMPs: MMP1, MMP12, and MMP13. S76 was the top hit for all three MMMPs, but additional substrates ranked differently as top hits. (B) The three queried proteases belong to the same protease class, but only two of the top hits were shared between the three proteases. (C) To determine the robustness of the rankings output by PAA, a new *in vitro* screen of MMP2, 12, and 13 was run against 6 of the "top hit" substrates and two other substrates that did not make this list (S19 and S22). (D) The cleavage efficiencies of the 8 selected substrates by the 3 MMPs were ranked relative to one another. (E-G) To visualize differences between the empirical vs. predicted rankings, the highest predicted rank of each substrate from the database query was plotted against the empirical rank of the substrate in the independent de novo screen. A positive correlation corresponds to agreement between the predicted rankings and the empirical results.

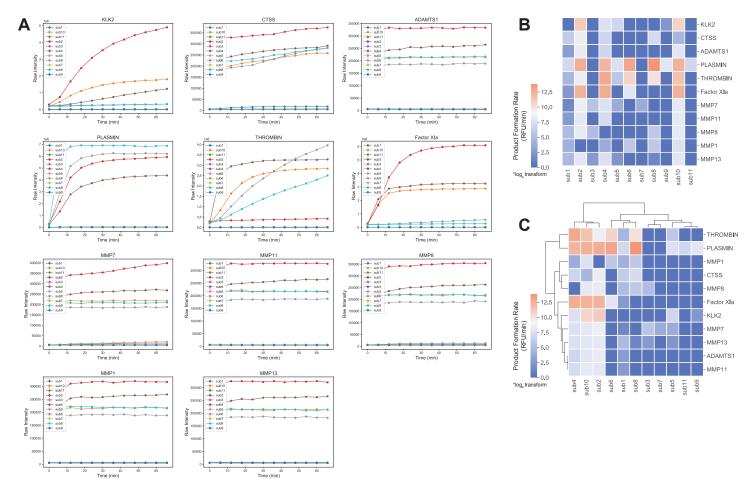


Figure S2: **PAA** enables faithful, automated analysis of protease activity data from the literature. Functions from PAA's KineticDataset were used to analyze kinetic substrate cleavage data from a protease-substrate *in vitro* screen from Holt and colleagues. ³² (**A**) PAA generates kinetic curves of raw fluorescence intensities as shown, for individual proteases screened against 11 substrates. Product formation rates are expressed in relative fluorescence units per unit time (RFU/min). (**B**) Heat map of product formation rates, calculated from kinetic activity curves in (A) and automatically generated by PAA. (**C**) PAA enables hierarchical clustering across proteases and substrates, rendering further insights into the relative cleavage similarities between proteases and substrates screened.